

Name	Ruttana Pachanon
Laboratory	Bioresources
Year (Grade)	D2
Place of practice	Faculty of Public Health, Thammasat University, Pathum Thani Province, Thailand
Period of practice	November 7 <sup>th</sup> 2017 – December 15 <sup>th</sup> 2017
Purpose	Surveillance of <i>Escherichia coli</i> and <i>Salmonella</i> isolated from swine farms and pork in Central region of Thailand

Summary of activities (about 800 words, provide photos, tables and figures that clearly show the activities during the period)

### Background and purpose

Foodborne diseases caused by *E. coli* and *Salmonella* have been recognized as a major concern for public health. These bacteria can be transmitted from animal reservoirs to humans by consuming contaminated water/food and through direct contact with the animals. Swine is a one of a food-producing animal. Moreover, the antimicrobial agents have been frequently used in swine production for therapeutic, growth promotion purposes and preventing the diseases, especially in the development of multidrug-resistant bacteria. The previous study in Thailand found that 68-99% of drug resistance phenotypes from *E. coli* and *Salmonella* isolated in swine (Changkaew et al., 2015, Chotinun et al., 2014). It suggested that there was a high incidence of antimicrobial resistance (AMR). Therefore, we need to determine the prevalence of *E. coli* and *Salmonella* in swine and their antimicrobial resistance to provide information for the nationwide surveillance of AMR.

**My activity at Faculty of Public Health, Thammasat University, Thailand  
during 7 November -15 December 2017**

**November 7<sup>th</sup> - 10<sup>th</sup> 2017:** I met the Dean, accompanying supervisor (Prof. Dr. Orasa Suthienkul) and investigator (Dr. Kanjana Changkaew) and then laboratory tour at Faculty of Public Health, Thammasat University. I have received a lecture about overview of conventional culture method for isolation and identification of *E. coli* and Nontyphoidal *Salmonella*. In the next day, culture media were prepared for isolation and identification of *E. coli* and *Salmonella*.

**November 13<sup>th</sup> - 24<sup>th</sup> 2017:** Culture media were prepared for isolation and identification of *E. coli* and *Salmonella*. The *E. coli* and *Salmonella* were isolated from 40 feces samples at Suphanburi province located in Central region of Thailand by conventional method.



**November 27<sup>th</sup> - 28<sup>th</sup> 2017:** Culture media were prepared for isolation and identification of *E. coli* and *Salmonella*.

**November 29<sup>th</sup> 2017:** Sample collection at Pathumthani market, Thailand

1. Collected pork and cutting board from 6 shops





Pork about 300 g

Pork on the surface of cutting board



## 2. Questionnaire survey by investigators



**November 29<sup>th</sup> to December 3<sup>th</sup> 2017:**

### **Sample preparation and enrichment procedure**

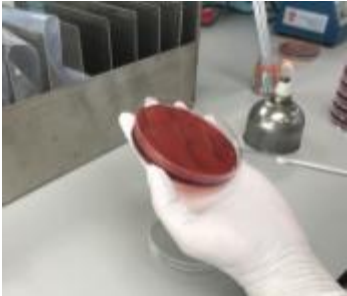

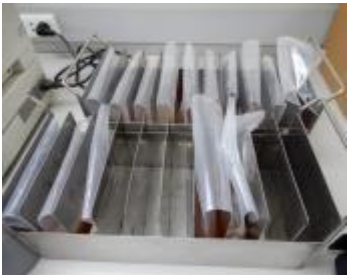


1. Prepare 300 g of pork sample and pork on the surface of cutting board



2. Weigh 25 g of pork into 225 ml of brain heart infusion (BHI) broth for *E. coli* and 225 ml of buffered peptone water (BPW) for *Salmonella* and stomach briefly as necessary.



3. Isolation and identification of *E. coli* and *Salmonella* by conventional culture method

<i>E. coli</i>	<i>Salmonella</i>
<ol style="list-style-type: none"> <li>1. Isolation of <i>E. coli</i> <ul style="list-style-type: none"> <li>● Direct method</li> </ul> </li> </ol>	<ol style="list-style-type: none"> <li>1. Isolation of <i>salmonella</i> <ul style="list-style-type: none"> <li>● Enrichment method</li> </ul> </li> </ol>
 <p>Streaked the sample onto Macconkey agar</p>	 <p>Swab the pork into 10 ml of buffered peptone water and incubated the sample at 37C°, overnight</p>
 <p>Incubated the sample at 37C°, overnight</p>	 <p>Rappaport-Vassiloadis (RV) enrichment broth</p>
<ul style="list-style-type: none"> <li>● Enrichment method</li> </ul>	



---

2. Identification of *E. coli* by biochemical tests



Picked up of pink colonies on Macconkey agar  
(approximately 1-5 colonies/sample)

↓  
**IMViC tests**



3. Interpretation of the result of biochemical tests

2. Identification of *salmonella* by biochemical tests



Positive                      Negative  
on Modified Semisolid Rappaport Vassiliadis  
(MSRV) agar



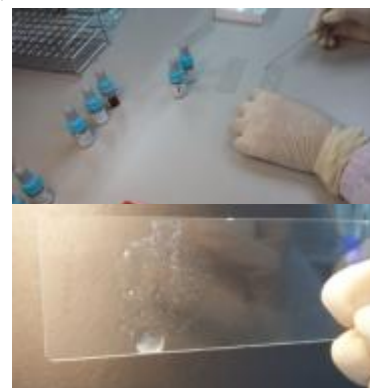
Picked up of colorless colony and HS<sub>2</sub> in the center  
on Xylose Lysine Deoxycholate agar  
(approximately 1-5 colonies/sample)

↓  
**TSI and LIM tests**



3. Interpretation of the result of biochemical tests

4. Serological test for serotyping of *Salmonella* spp.



Positive for serological test

---

### Other identification (methicillin resistant *S. aureus*)

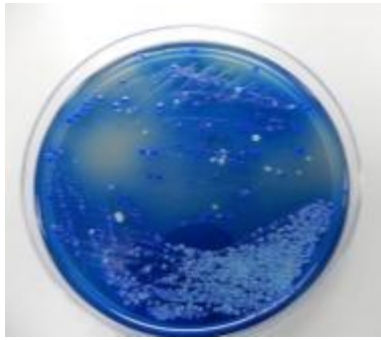
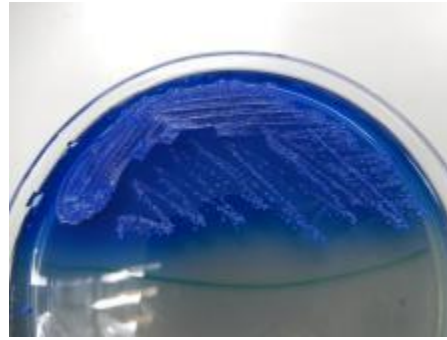


Plate from sample



Positive for *S. aureus* (Reference)

Blue colonies on oxacillin resistance screening agar base (ORSAB) plates



Positive: clotting

Negative

Coagulase test for *S. aureus*



Catalase test for *S. aureus*

**December 4<sup>th</sup> 2017:** Attended a distinguished lecture by Prof. Dr. Hiroshi Kida

**December 6<sup>th</sup> – 7<sup>th</sup> 2017:**

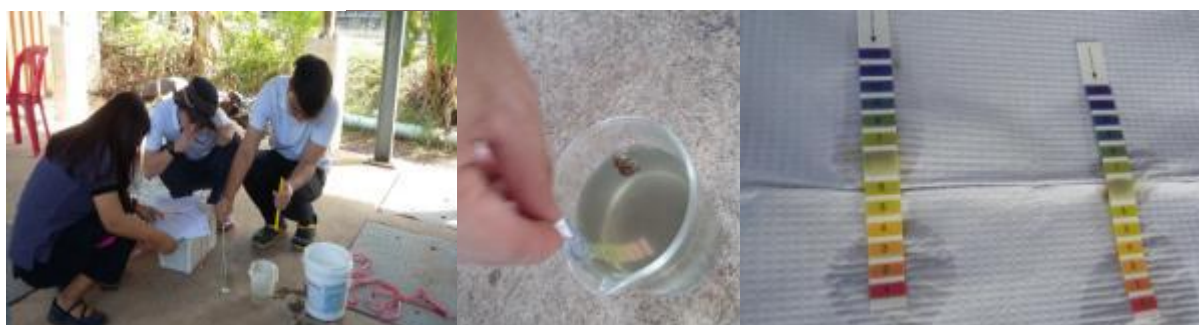
- Isolation and identification of *Salmonella* spp.
- Attended Mr. Kentaro Koide's presentation

**December 8<sup>th</sup> – 15<sup>th</sup> 2017:**

- Presentation of my research activity
- DNA extraction of the *E. coli* isolates by boiled method
- Testing PCR condition of *E. coli* reference strain

## Additional sample collection

1. Waste water in area Thammasat University (November 23<sup>th</sup> 2017)



2. Air sampling for bacteria and fungi at Faculty of Medicine (November 28<sup>th</sup> 2017)



## Results

- The prevalence of *E. coli* and *Salmonella* in swine from farms, pork and pork on surface of cutting board from markets as shown below;

Source	Total of samples	No. positive of <i>E. coli</i>		No. positive of <i>Salmonella</i>	
		Samples	Isolates	Samples	Isolates
<b>Farm</b> (Feces)	40	40	327	5	14
<b>Market</b>					
- Pork	6	6	41	4	7
- Pork on surface of cutting board	6	6	44	4	13

## Outcome

- I have learned more about field epidemiology for sample collection in the market, isolation and identification of *E.coli* and *Salmonella* spp. including methicillin resistant *S. aureus* (MRSA).
- I have more skill of the serological test for serogrouping of *Salmonella* spp. and also molecular technique for DNA extraction and testing PCR condition.
- However, *Salmonella* serovar will be confirmed by *Salmonella* and *Shigella* Center, Nonthaburi Thailand. And also the team (investigators at Faculty of Public Health, Thammasat University), they need to further determine the antimicrobial resistant pattern of *E.coli* and *Salmonella*. In the future, I may be obtained opportunity from the collaborative at Faculty of Public Health, Thammasat University, Thailand.

(Field Epidemiology • Collaborative Research) Evaluation by supervisor

Institution • Official title • Name	Prof. Yasuhiko Suzuki	印
Describe overall evaluation on the applicant's activity in overseas practice.		